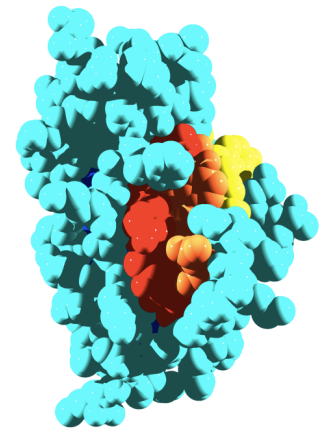
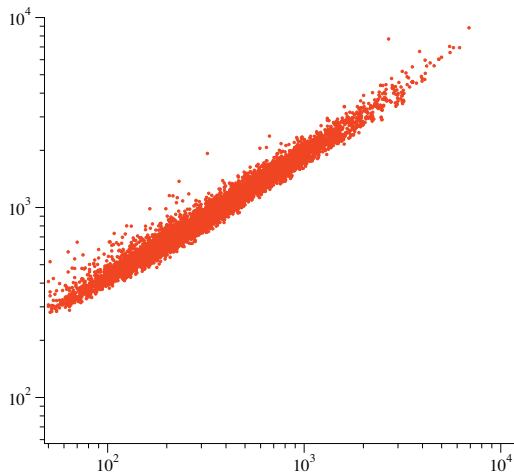


# Scaling Laws, the Golden Ratio, & the Small-Angle Scattering of Biomolecules

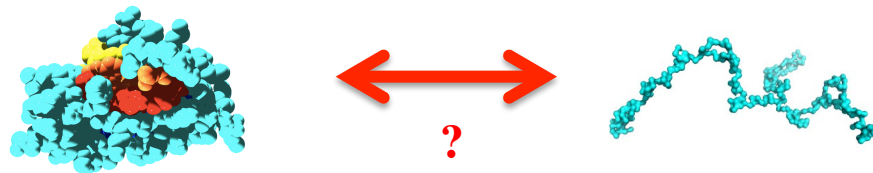


Max C. Watson

I) A fast method for calculating scattering intensities

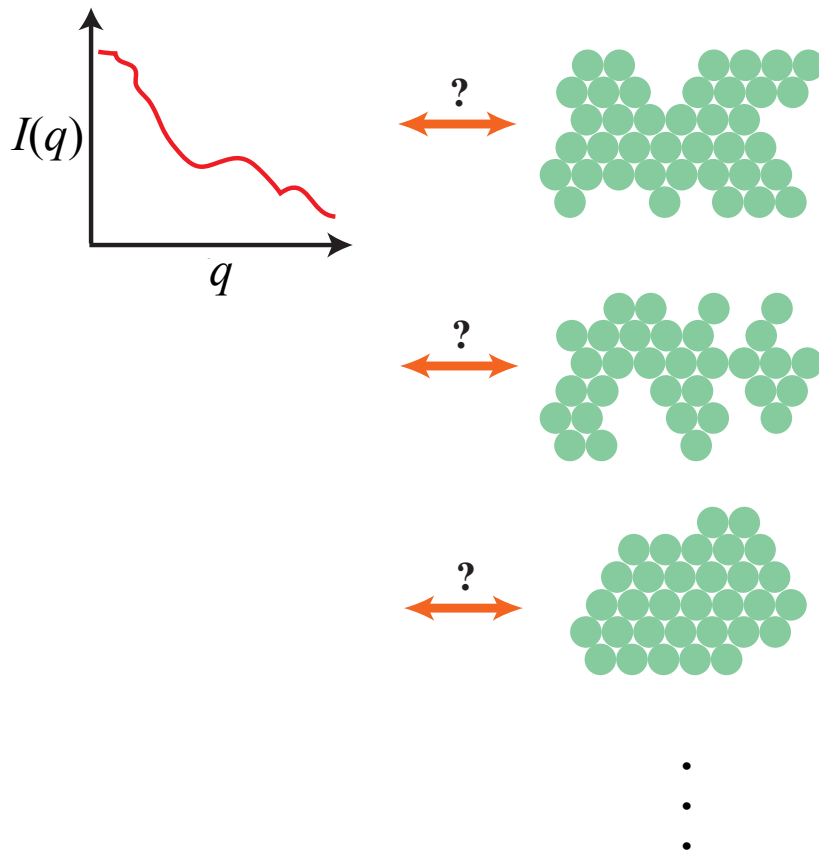


II) Scaling Laws & Molecular Disorder

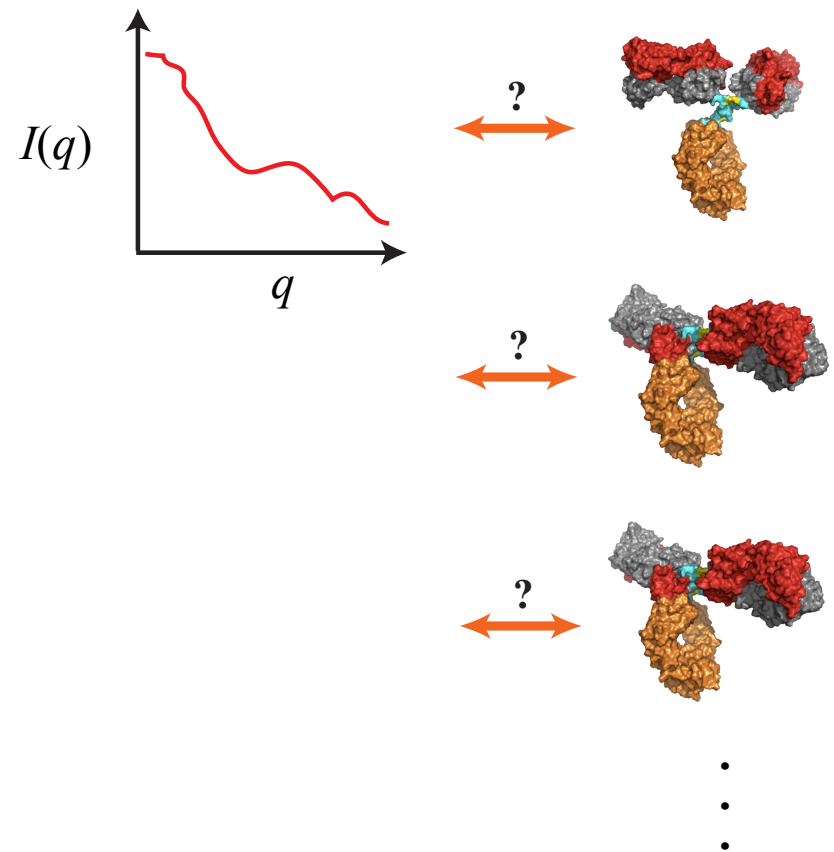


# Shape Determination

## Ab Initio Methods



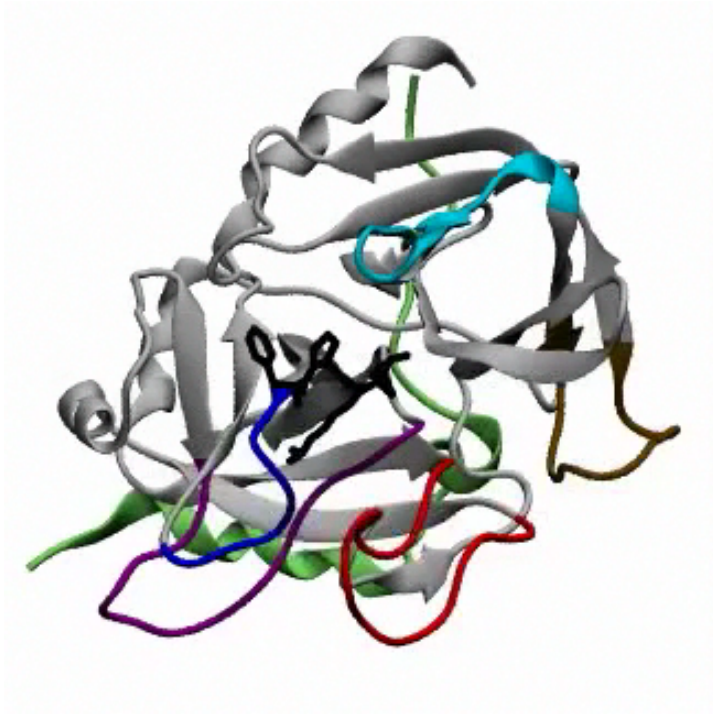
## Rigid Body Modeling



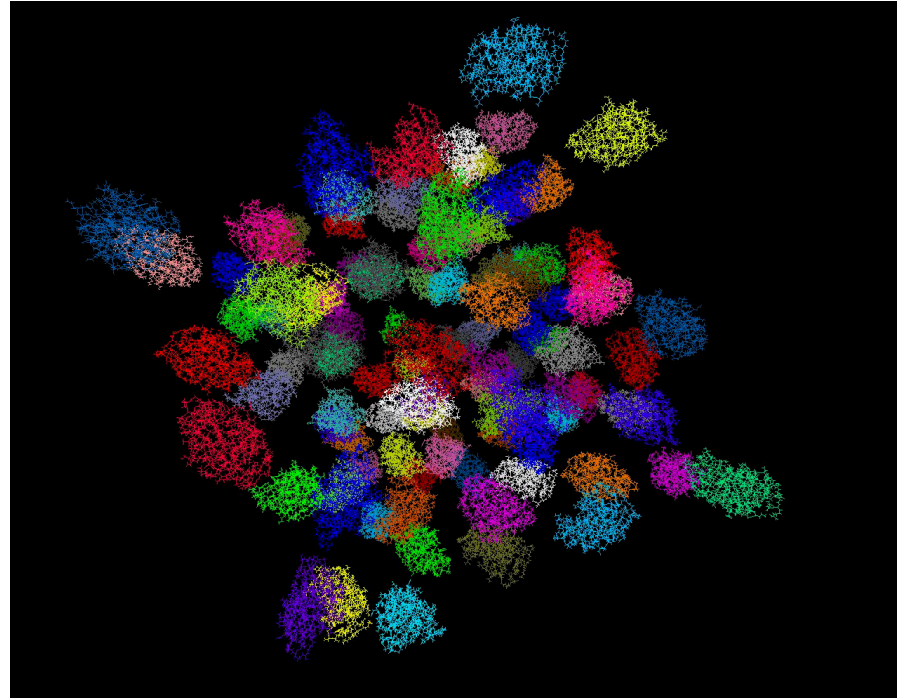
Chacon et al. 1998, Svergun 1999

Wall et al. 2000, Curtis et al. 2012

# Connection to Simulations



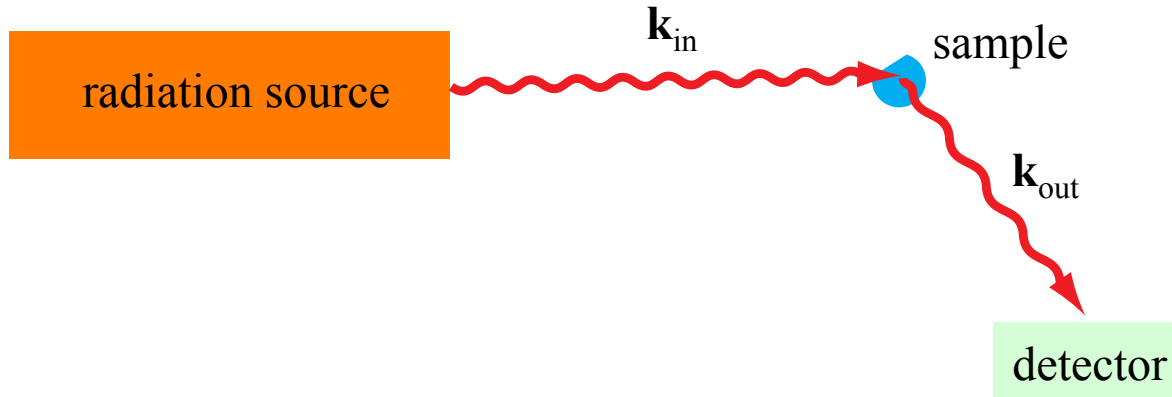
Single Protein  
Fuglestad et al. 2012



Concentrated Protein Solutions



# Scattering Fundamentals



amplitude

$$A(\mathbf{q}) = \sum_j^N b_j e^{-i\mathbf{q} \cdot \mathbf{r}_j}$$

$\mathbf{q} = \mathbf{k}_{\text{out}} - \mathbf{k}_{\text{in}}$  scattering vector  
 $\mathbf{r}_j$  position of atom  $j$   
 $b_j$  scattering length of atom  $j$   
 $N$  number of atoms

scattering  
intensity (flux)

$$I(\mathbf{q}) = |A(\mathbf{q})|^2$$

experimental  
observable

$$I(q) = \langle I(\mathbf{q}) \rangle_{\text{all directions}}$$

# Easier Said Than Done

$$I(q) = \langle I(\mathbf{q}) \rangle_{\text{all directions}}$$

Exact Result: 
$$I(q) = \sum_j^N \sum_k^N b_j b_k \frac{\sin(q|\mathbf{r}_j - \mathbf{r}_k|)}{q|\mathbf{r}_j - \mathbf{r}_k|} \quad (\text{Debye 1915})$$

Summing over all pairs is an  $O(N^2)$  calculation

For one protein,  $N \sim 10^3 - 10^6$

Even worse for:    multiple proteins  
                         flexible domains  
                         shape determination

# A Simple Alternative

$$I(q) = \langle I(\mathbf{q}) \rangle_{\text{all directions}}$$

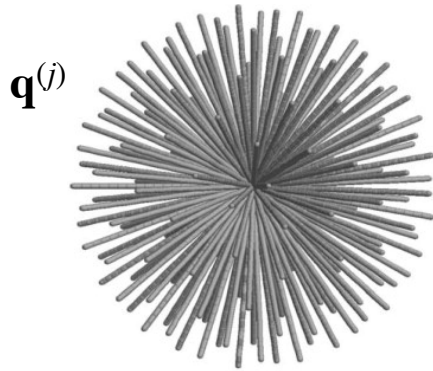
$\mathbf{q}$ : scattering vector

$I(\mathbf{q})$ :  $\mathbf{q}$ -dependent scattering intensity

$I(q)$ : measured scattering intensity

Numerically calculating  $I[\mathbf{q}]$  at a given  $\mathbf{q}$  scales as  $O[N]$  ( $N$  = number of atoms)

To get  $I(q)$ , just average over  $I[\mathbf{q}]$  for many  $\mathbf{q}$ 's :

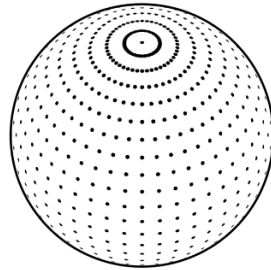


$$I(q) \approx \frac{1}{n} \left[ \sum_j I(\mathbf{q}^{(j)}) \right]$$

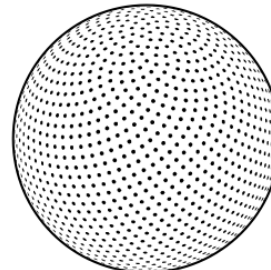
Calculating  $I(\mathbf{q})$  then scales as  $O[nN]$

# Generating a Quasi-Spherical Lattice

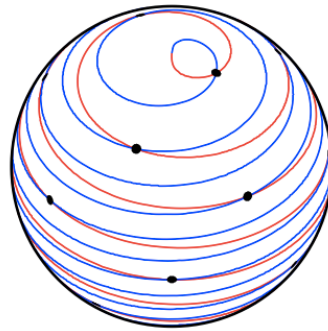
Bad!



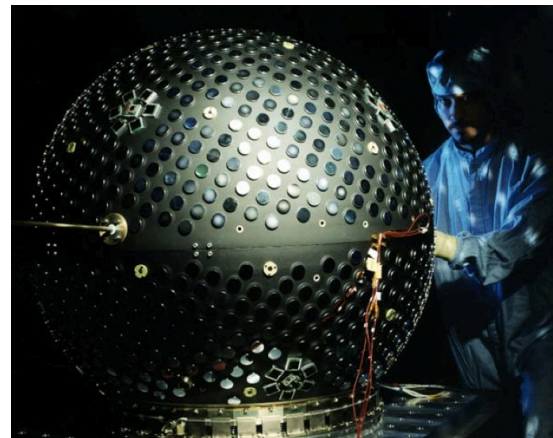
Good

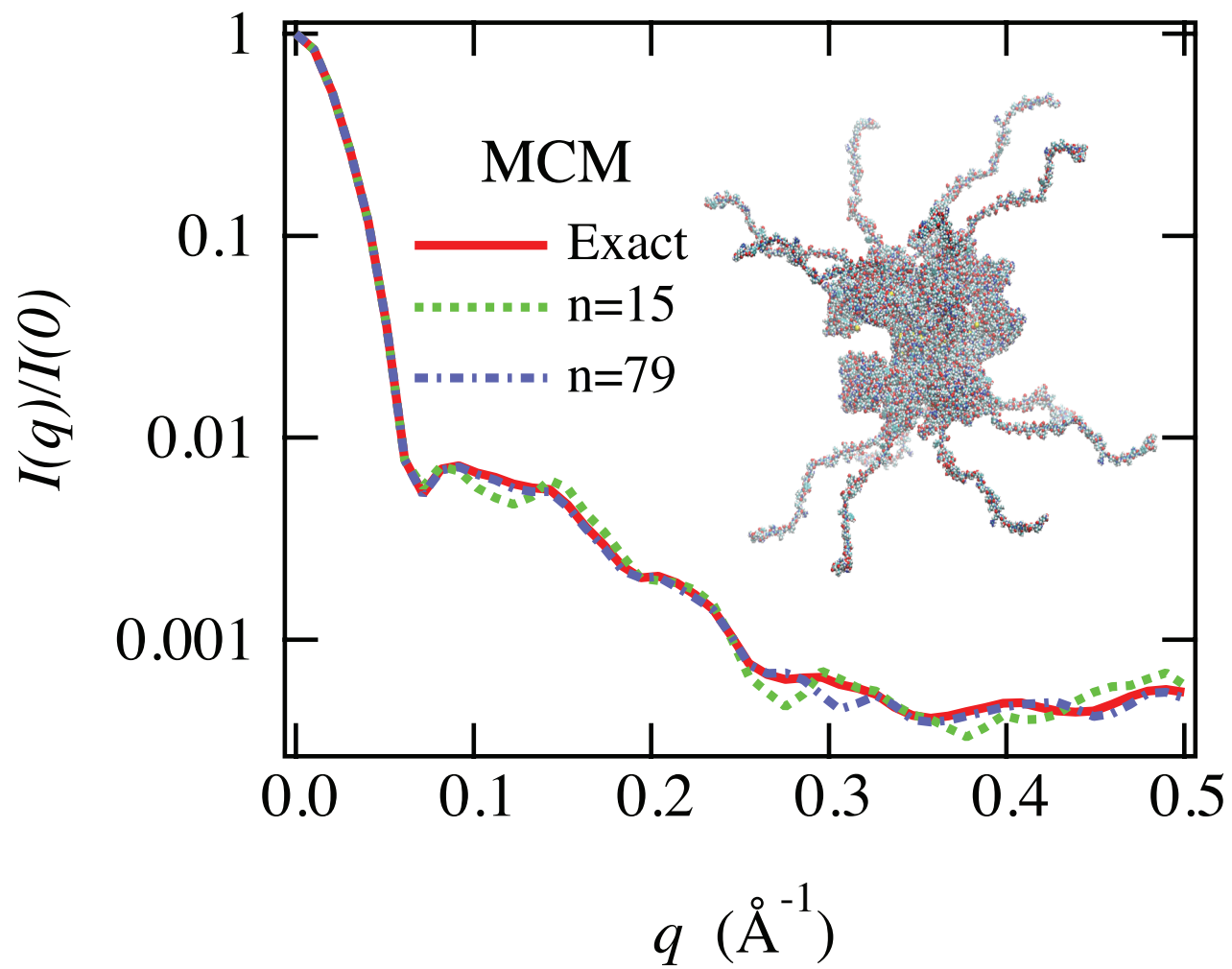


Fibonacci Lattice Built  
Using Golden Ratio:  
 $\Phi = 1.618\dots$



(González 2010)

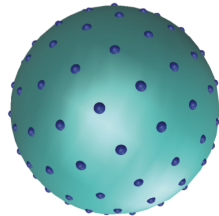




(Krueger et al, 2011)

# Golden Vector Method

Step 1: generate  $n$  scattering vectors  $\mathbf{q}^{(j)}$  on quasi-spherical lattice using golden ratio



Step 2: calculate  $I[\mathbf{q}^{(j)}]$  for each  $\mathbf{q}^{(j)}$

Step 3: average over all  $I[\mathbf{q}^{(j)}]$ : 
$$I(q) \approx \frac{1}{n} \left( \sum_{\mathbf{q}^{(j)} \in \text{lattice}} I[\mathbf{q}^{(j)}] \right)$$

- Speed scales as  $O[nN]$
- For given level of accuracy, 2-8 times faster than Spherical Harmonic Method (CRYSON)
- Watson and Curtis, *Journal of Applied Crystallography*, 2013



# Summary

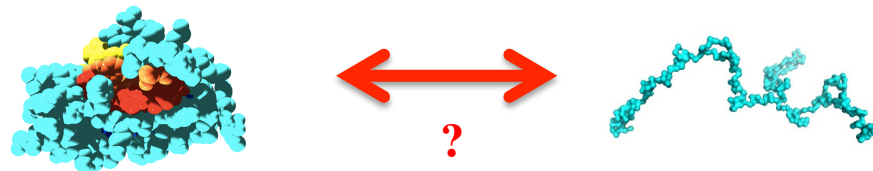
- Golden Vector Method: simple yet powerful
  - good for any collection of atoms
  - easy to customize
- Ongoing Work at NCNR:
  - analysis of polymer simulations (Mike Hore)
  - incorporate solvent effects and web server (Hailang Zhang and Joseph Curtis)



I) A fast method for calculating scattering intensities

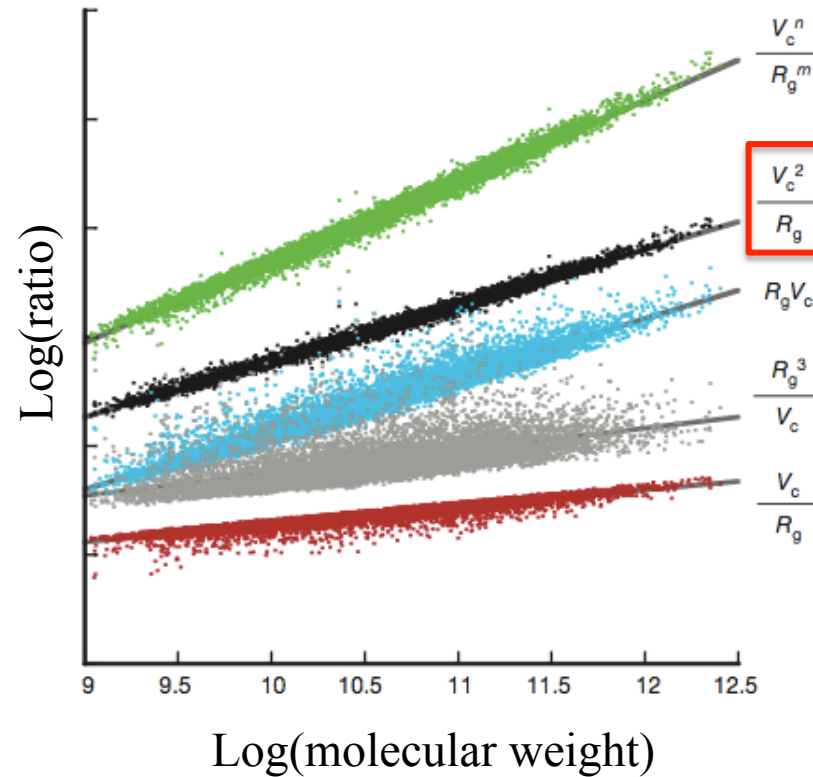


II) Scaling Laws & Molecular Disorder



# A Curious Result for Compact Proteins:

$$V_c \equiv \left( \int_{\text{all } q} \frac{I(q)}{I(0)} q \, dq \right)^{-1} \quad R_g = \text{radius of gyration}$$



Rambo & Tainer  
2013

$$\text{molecular weight} \propto \frac{V_c^2}{R_g}$$

# What is $V_c$ really?

$$V_c(q_m) \equiv \left( \int_0^{q_m} \frac{I(q)}{I(0)} q \, dq \right)^{-1} \quad q_m = \text{adjustable parameter}$$

Properties well known for  $q_m \rightarrow \infty$ , but what about finite  $q_m$ ?

It can be shown that:

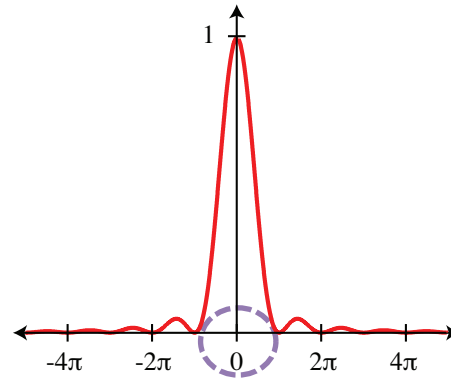
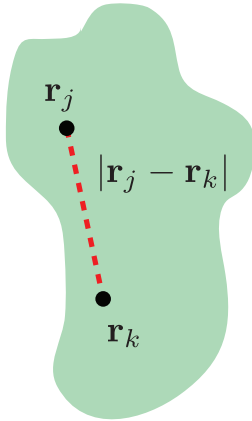
$$V_c(q_m) = \frac{2I(0)}{q_m^2 J(q_m)}$$

$$J(q_m) = \sum_j^N \sum_k^N b_j b_k \operatorname{sinc}^2(q_m |\mathbf{r}_j - \mathbf{r}_k|/2) \quad \text{and} \quad \operatorname{sinc}(x) \equiv \frac{\sin(x)}{x}$$

$\mathbf{r}_j$	position of atom $j$
$b_j$	scattering length of atom $j$
$N$	number of atoms

$$J(q_m) = \sum_j^N \sum_k^N b_j b_k \operatorname{sinc}^2(q_m |\mathbf{r}_j - \mathbf{r}_k|/2) \quad j, k \text{ each run over all atoms}$$

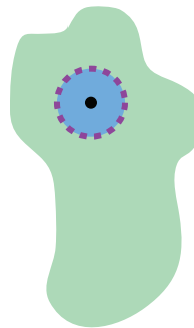
$$\operatorname{sinc}^2(x)$$



$q_m$  describes an effective probe size

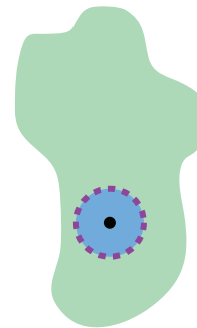


$$J(q_m) =$$



$k=1$

+

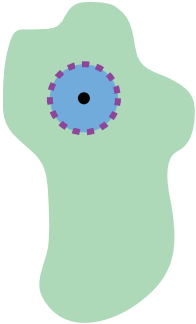
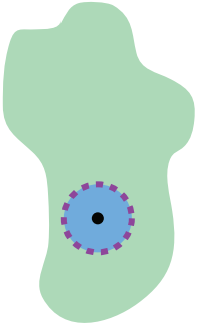


$k=2$

+ ...

Compact Molecules:

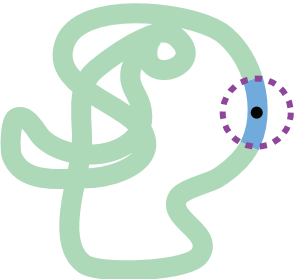
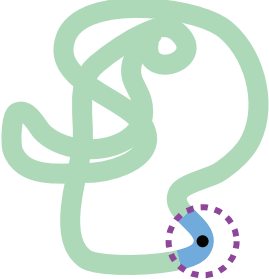
$$J(q_m) =$$


+

+ ...

$k=1$ 
 $k=2$

Disordered molecules:

$$J(q_m) =$$


+

+ ...

$k=1$ 
 $k=2$

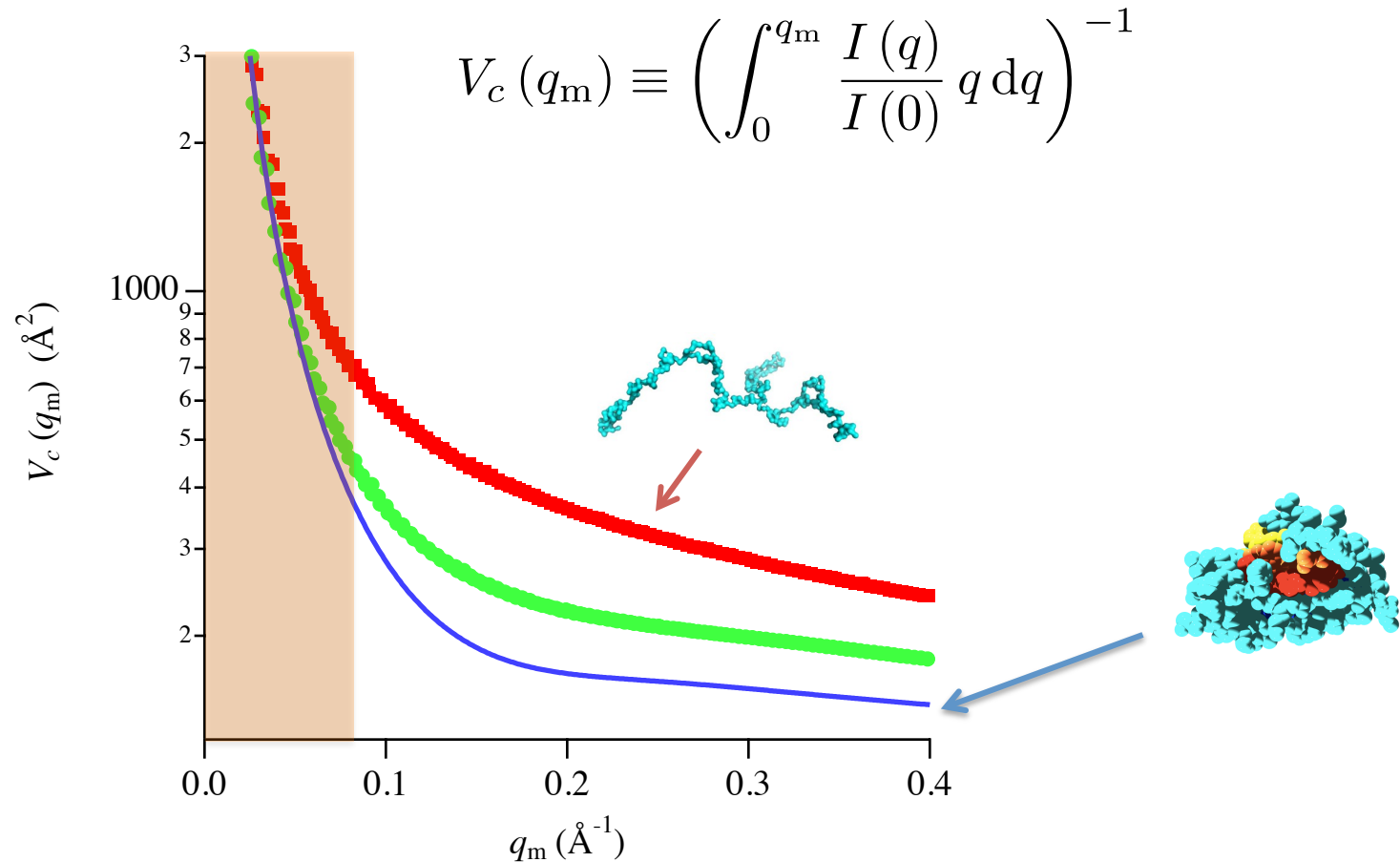
smaller than  $J(q_m)$   
for compact molecules

$$V_c(q_m) = \frac{2I(0)}{q_m^2 J(q_m)}$$

$V_c(q_m)$  is \*larger\* for disordered molecules, for a given number of atoms



# $V_c$ probes molecular disorder



information about disorder is even contained at very low  $q_m$

# Scaling Laws for $V_c$

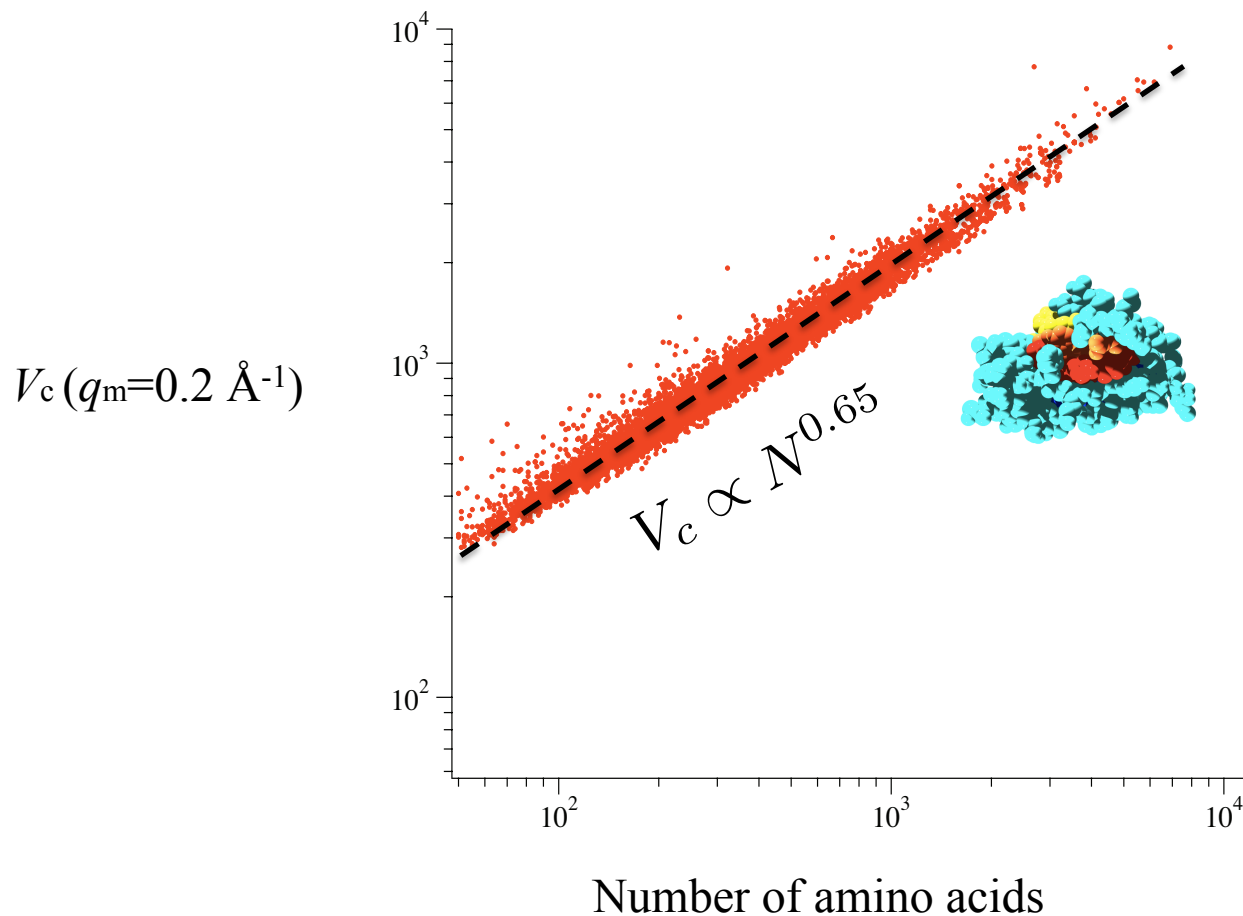
$$q_m^{SAS} = (0.2 - 0.5) \text{ \AA}^{-1} \quad \text{Upper limit for Small-Angle Scattering (SAS)}$$

Calculate  $V_c (q_m^{SAS})$  :

Guinier Approximation:  $\frac{I(q)}{I(0)} \approx \text{Exp} \left( -\frac{R_g^2 q^2}{3} \right) \quad (qR_g \lesssim 1)$

For compact proteins:  $\left\{ \begin{array}{l} V_c (q_m^{SAS}) \approx \frac{2}{3} R_g^2 \\ R_g \propto N^{1/3} \quad N = \text{number of atoms} \\ V_c (q_m^{SAS}) \propto N^{2/3} \end{array} \right.$

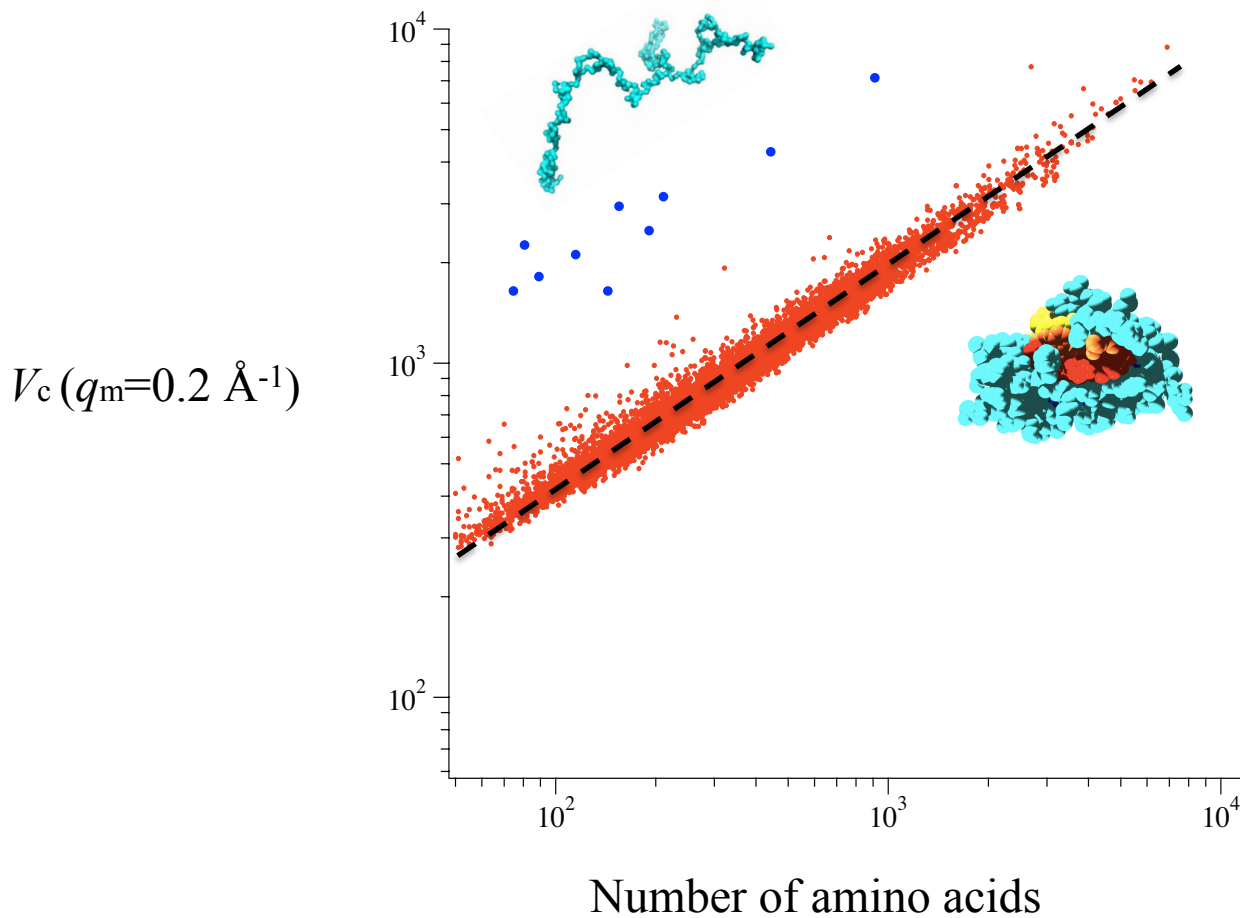
# Theory Agrees with Compact Protein Data



$V_c \propto N^{2/3}$  explains Rambo and Tainer finding:

$$\text{molecular weight} \propto \frac{V_c^2}{R_g}$$

# Disordered Proteins Have a Larger $V_c$

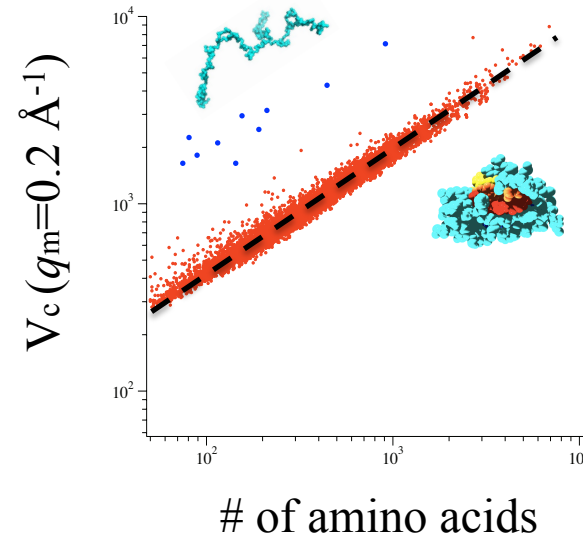
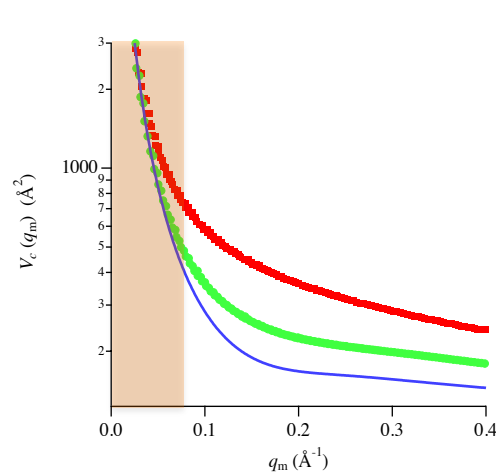


Position in plot can be used to quantify level of disorder

# Summary of $V_c$

$$V_c(q_m) \equiv \left( \int_0^{q_m} \frac{I(q)}{I(0)} q \, dq \right)^{-1}$$

- Concentration independent, relative scale, easy to calculate
- Useful for comparing two molecules with roughly same # of atoms in a model-free way
- Contains good information even at low  $q_m$



- Can estimate molecular weight (compact)
- Can be applied to other molecules too (e.g. RNA, polymers...)

# Acknowledgements

- Joseph Curtis, Susan Krueger, Michi Nagao, Nick Clark
- Dan Neumann and Rob Dimeo
- National Research Council

